Epithelial Cytokines: Connecting the Dots in Airway Diseases

This industry-sponsored symposium took place during the American Thoracic Society (ATS) International Conference held in San Diego, California, USA, from May 20th-22nd, 2024.

Speakers:	Rohit Katial, ¹ Andrew White, ² Igor Barjaktarevic ³
	 Center for Clinical Immunology, National Jewish Health, University of Colorado, Denver, USA Division of Allergy, Asthma and Immunology, Scripps Clinic, San Diego, California, USA Division of Pulmonary and Critical Care Medicine, David Geffen School of Medicine, University of California, Los Angeles (UCLA), USA
Disclosure:	Katial has received consultancy fees and speaker honoraria fees from AstraZeneca, Amgen, Sanofi/Regeneron, and Grifols; consultancy fees from GSK; and speaker honoraria from Pharming. White has received speaker honoraria from AstraZeneca, Amgen, Regeneron/Sanofi, Blueprint Pharmaceuticals, Genentech, Optinose, and GSK; has been a member of the advisory board for Sanofi, GSK, AstraZeneca, and Optinose; and has provided research support for Regeneron. Barjaktarevic has received grant support from NHLBI, ALA, COPD Foundation, Alpha-1 Foundation, Theravance/Viatris, Aerogen, Takeda, GE Healthcare, and Amgen; and has received consulting fees from Verona Pharma, Sanofi/Regeneron, Genentech, AstraZeneca, Inhibrx, Aerogen, Takeda, and Grifols.
Acknowledgements:	Medical writing assistance was provided by Eleanor Roberts, Beeline Science Communications, Ltd, UK. Patients gave informed, written consent for video recordings and to the use of the video recordings.
Disclaimer:	This article is for disease state education, and is not meant to imply efficacy or safety of any AstraZeneca products or other medications.
Keywords:	Airway epithelium, cytokines, hyperresponsiveness, immune cells, inflammation, nasal polyps remodeling, unmet needs.
Citation:	Respir AMJ. 2025;3[Suppl 1]:2-9. https://doi.org/10.33590/respiramj/IXJL9681



2

Summary

Understanding the role of the epithelial cytokines thymic stromal lymphopoietin (TSLP), IL-25, and IL-33 in inflammatory airway diseases such as severe asthma (SA), chronic rhinosinusitis with nasal polyps (CRSwNP), and chronic obstructive pulmonary disease (COPD) could lead to new insights into the pathophysiology of airway diseases. The airway epithelium plays a critical role in these diseases, with TSLP,

IL-25, and IL-33 released following exposure to disease triggers, such as allergens, pollutants, and pathogens, which can exacerbate these conditions. These cytokines can also be released from both immune and nonimmune cells and can act on a broad array of target cells. In asthma, the release of TSLP, IL-25, and IL-33 following exposure to a trigger has been linked to airway inflammation and hyperresponsiveness. Epithelial disruption in CRSwNP can drive inflammation, impair mucociliary clearance, and increase permeability and mucus production. Epithelial cytokine elevation is especially prominent in eosinophilic CRSwNP, where it is positively correlated with symptoms and underlying pathophysiology. Conversely, causative mechanisms in COPD are poorly understood, and management is sometimes inadequate. However, similar to asthma, IL-33 and TSLP are increased in patients with COPD, indicating that they may also have a role in the pathophysiology of this disease. Significant challenges remain regarding understanding of SA, CRSwNP, and COPD, especially in patients where symptoms are uncontrolled. Therefore, elucidating the role of TSLP, IL-25, and IL-33 in these conditions may have the potential to offer new insight into these conditions.

Introduction

Understanding of the airway epithelium initially focused on its role as a protective barrier and environmental sensor. However, it is now known that numerous, and in some individuals, multiple, environmental triggers can initiate epithelial-driven innate and adaptive immune responses.¹ These can lead to the inflammatory changes seen in acute and chronic obstructive airway diseases, structural changes, and airway remodeling (Figure 1).²⁻⁵ As such, the airway epithelium is becoming a focus for elucidating the pathophysiology of inflammatory conditions such as SA, CRSwNP, and COPD.

While descriptions of upper and lower airway diseases have been noted for centuries, elucidation of their pathogenesis has only occurred in the last 150 years, with more advanced descriptions of associated inflammatory signaling stemming from the mid-1980s.⁶⁻⁸ One such development has been additional understanding of the role of three airway epithelium-produced proinflammatory cytokines with roles in SA, CRSwNP, and COPD: TSLP, IL-25, and IL-33.9 While these diseases have their own manifestations, findings regarding concurrence of global prevalence, similar cell and tissue type representation, and shared phenotypes support a unified airway model regarding their pathogenesis,¹⁰ which may involve these epithelial cytokines.¹¹

Human TSLP was first identified in 2001,¹² with its role in activation of a key cell type involved in airway diseases, Type 2 innate lymphoid cells (ILC2), not described until 2016.¹³ IL-25 was also first described in 2001,¹⁴ with description of IL-33 in human endothelial cells dating to 2003.¹⁵ These epithelial cytokines can activate inflammatory responses in both distinct and overlapping ways, and may have implications in the underlying inflammatory processes in SA, CRSwNP, and COPD (Figure 1).¹⁶

While the airway epithelium is the primary source of TSLP, IL-25, and IL-33, they can also be produced by a range of both immune cells (Figure 1) and non-immune cells, such as stromal cells, fibroblasts, and smooth muscle cells. These epithelial cytokines can also target a wide array of cells, leading to complex, bi-directional cell-cell interactions.¹⁶⁻²⁰ Following activation of inflammatory cells by TSLP, IL-25, and IL-33, proinflammatory cytokines expressed by the affected cells include IL-4, IL-5, IL-9, IL-13, and granulocyte macrophage colony stimulating factor.^{2,16-20} These mediators can contribute to upper and lower airway disease development and pathology, including inflammation, airway hyperresponsiveness, mucus production, and potentially, airway remodeling.²⁰⁻²²

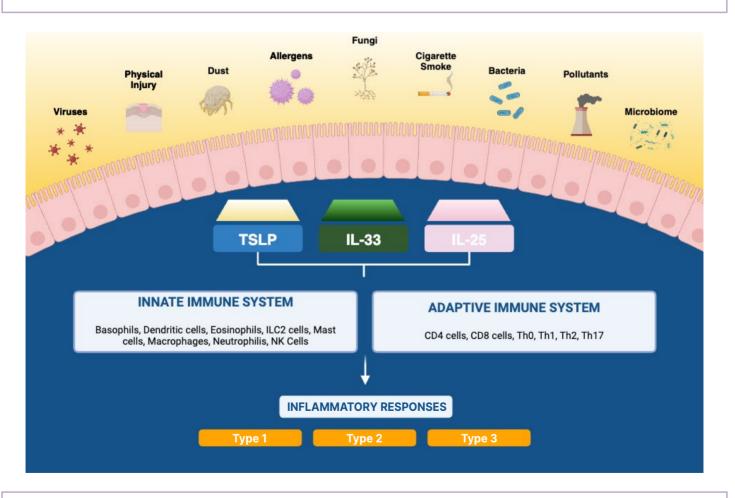
Severe Asthma



Around 262 million people worldwide are affected by asthma, a condition involving excessive bronchoconstriction

due to airway inflammation, airway hyperresponsiveness (which can occur regardless of the level of airway inflammation), and, in some patients, reduced lung function.²³ Asthma is classed as severe in around 5–12% of people with the condition.²⁴ One Canadabased study found that, compared to nonsevere asthma, SA accounts for at least half of all asthma-related excess healthcare costs, especially when it is comorbid with other medical conditions.²⁵ Patients with severe asthma can have multiple asthma triggers (Figure 1). For example, in a study by Chipps and colleagues,¹ patients had on average eight different triggers. This study also found that as trigger number increases, disease control can lower, asthma exacerbations and hospitalizations can increase, work productivity can decrease, and health-related quality of life can worsen.¹ Most patients with SA show activation of multiple inflammatory pathways and their respective biomarkers. For example, a study of patients from the International Severe Asthma Registry (ISAR) found that, of 1,175 adults with SA, nearly all (88%) had between 1-3 biomarkers for asthma, as evidenced by allergen-specific





ILC2: Type 2 innate lymphoid cell; NK: natural killer; Th: T-helper; TSLP: thymic stromal lymphopoietin.

serum IgE positivity \geq 75 kU/L, fractional exhaled nitric oxide levels \geq 25 parts per billion, or blood eosinophils \geq 300 cells/mL).²⁶

Similarly, a post hoc analysis of multiple Phase IIb/III trials found that, of 740 participants with SA with blood eosinophils \geq 150 cells/mL, nearly 80% had allergic asthma, and nearly 50% had fractional exhaled nitric oxide levels \geq 25 parts per billion.²⁷

Release from the airway epithelium of TSLP, IL-25, and IL-33, and subsequent activation of multiple inflammatory pathways^{2,16,21} is linked to several hallmarks of SA, including airway hyperresponsiveness²⁸ and remodeling,²⁹ decreased lung function,³⁰ and increased asthma severity,^{30,31} exacerbation risk,³² and mucus production.^{17,33}

The epithelial cytokines IL-25, IL-33, and TSLP may play a role in initiation and persistence of airway inflammation, leading to manifestations of asthma. For example, allergen challenge can significantly increase the number of bronchial epithelium and submucosa cells expressing these epithelial cytokines,²⁸ and airway concentrations of TSLP and IL-33 are significantly increased in patients with asthma, particularly SA, compared with healthy controls.³⁰ Airway inflammation can contribute to lower lung function, and a significant inverse correlation has been shown between forced expiratory volume in 1 second and both TSLP and IL-33 levels in bronchoalveolar lavage fluid, and the number of bronchial epithelium and submucosa cells reactive for these cytokines.28,30

TSLP and IL-33 levels are also significantly increased in patients with asthma who use inhaled corticosteroids (with or without oral corticosteroids) compared with patients who do not use these treatments, indicating that levels of these epithelial cytokines increase as the need for medication increases.^{30,31} Additionally, higher serum TSLP levels (≥25 pg/mL) are associated with higher exacerbation risk in SA, suggesting that TSLP appears to be involved in the pathogenesis of asthma exacerbations.³² Indications that these cytokines have a role in downstream pathways associated with asthma induction and severity come from several investigations. For example, upregulation of OX40 ligand on dendritic cells by epithelium-derived TSLP can induce an interaction with OX40 on T cells, polarizing them to a Type 2 phenotype and leading to the release of IL-4, IL-5, and IL-13.^{2,16,22}

Levels of these inflammatory cytokines correlate significantly with lung function in asthma.³⁰ They are involved in SA-linked pathologies including nitric oxide induction, IgE production, goblet cell hyperplasia (goblet cell hyperplasia leads to mucus hypersecretion and may be involved in mucus plug formation), and smooth muscle reactivity, linked to bronchial hyperresponsiveness ^{2,21} Additionally, IL-5 is the primary growth factor for eosinophils, and upregulation by IL-5 promotes eosinophilic inflammation.^{2,22}

Chronic Rhinosinusitis with Nasal Polyps



As a gateway to the respiratory system, nasal airway epithelium is often the first point of contact for microorganisms, pollutants, and allergens.³ Although the

immune response launched by airway epithelial cells is important for defense, it can also drive inflammatory diseases. For example, in allergic rhinitis, allergen exposure drives IgE-mediated mucosal inflammation that can become chronic.³ Chronic rhinosinusitis, generally categorized into occurrence with or without nasal polyps, is diagnosed in patients with symptoms and evidence of sinus inflammation on nasal exam or imaging that has persisted for at least 12 weeks.³⁴

CRSwNP, which typically occurs between ages 40–60 years,³⁵ is estimated to globally affect up to 4.4% of the population, with a slightly higher prevalence in males.^{35,36}



Cardinal signs and symptoms of CRSwNP include nasal congestion, loss of sense of smell, and impaired sleep quality, in addition to facial pain and pressure and nasal discharge. Loss of the sense of smell is postulated to be the result of blockage of access, along with inflammation driven damage, to the olfactory tissue.^{34,35}

Based on a USA-based study, compared with patients without chronic rhinosinusitis, patients with CRSwNP are estimated to visit an emergency department 1.5 times as often, a healthcare professional 2.0 times as often, and have incremental healthcare costs 2.5 times greater.³⁷

Troublesome symptoms of CRSwNP, such as nasal congestion and loss of the sense of smell,^{38,39} can have not only physical but also mental and social impacts on healthrelated quality of life.³⁹ Despite such findings, patients with CRSwNP express frustration due to perceived underestimation of the impact of the disease.³⁸

Around two-thirds of patients with CRSwNP also have asthma, with this comorbidity leading to a greater burden of disease, including more difficulty in gaining asthma control and a higher likelihood of experiencing polyp recurrence and steroid dependence.¹¹ The idea that CRSwNP and asthma share common underlying inflammatory pathways is supported by studies showing that allergen challenges in either upper or lower airway alone can lead to subsequent presence of inflammatory mediators in both locations. This may be linked to expression in both diseases of epithelium cytokines,⁴⁰ supporting the unified airway model.¹⁰

As in SA, in CRSwNP, the epithelium is not just a passive barrier, with damage only caused by immune system cells, but is an intricate part of the pathophysiology of the disease. Damage to the sinus epithelium can trigger cell proliferation and transformation of epithelial cells to a mesenchymal form, leading to inflammatory processes and remodeling.⁴¹ In addition, upregulation of mast cell and basophil degranulation results in vasodilation, with leakage of plasma containing fibrin and edema. This is followed by accumulation of cross-linked fibrin, which forms the bulk of the nasal polyp, forming a dense mesh capable of trapping plasma proteins and water, leading to edema and remodeling.⁴²

Three inflammatory endotypes of CRSwNP have been described, which can be mixed in many patients, or not present in a minority.⁴³⁻⁴⁵ Type 1 is characterized by the release of interferon- γ from ILC1, Th1 cells, cytotoxic T cells, and natural killer cells.⁴⁴ Dominant clinical features include headache and facial pain.⁴⁵ Cytokines released in Type 3 CRSwNP include IL-17 and IL-22 from ILC3s, Th17 cells, and neutrophils.⁴⁴ Key features of this endotype include eye itching and purulent rhinorrhea.⁴³⁻⁴⁵

However, most patients with CRSwNP are Type 2, characterized by anosmia, asthma comorbidity, and high clinical burden.⁴³⁻⁴⁵ In this endotype, upregulation of TSLP, IL-25, and IL-3342 can lead to the release of IL-4, IL-5, and IL-13 from eosinophils, and Th2 cells, basophils, and ILC2s.^{42,44} Higher levels of these epithelial cytokines or markers for them have been found in nasal polyp tissue of patients with CRSwNP compared with uncinate process or turbinate tissue of controls.⁴⁶⁻⁴⁸ Studies also point to a role of some of these epithelial cytokines in conversion of ILC2 cells to a corticosteroid-resistant subtype.⁴⁹ This is borne out by findings that TSLP levels are significantly increased in nasal epithelial cells in patients who have corticosteroidresistant nasal polyps.⁵⁰

The epithelial cytokines are particularly increased in eosinophilic CRSwNP (eCRSwNP) compared with controls and with patients with non-eCRSwNP.⁵¹ There are also correlations between disease severity in patients with eCRSwNP and expression of TSLP, the TSLP receptor, and the membrane-bound IL-33 receptor ST2L,

based on patient ratings of symptoms and CT scan.⁵² Similar to SA, these findings suggest an intimate role of the epithelial cytokines in CRSwNP pathology.^{48,52}

Chronic Obstructive Pulmonary Disease



Globally, an estimated 392 million people aged 30–79 years live with COPD,⁵³ with numbers,⁵⁴ along with

healthcare-related costs,55 predicted to rise over the coming decades. The number of adults living in the USA with COPD is currently around 14.2 million, with little recent change in prevalence over the time period 2011 to 2021.56 In 2021, COPD was the sixth leading cause of death in the USA, with fatality numbers at nearly 140,000.⁵⁷ Management options for COPD may not sufficiently target inflammatory pathways that contribute to this condition.⁴ This is evidenced by a number of findings, including that of a study that showed that half of hospital-treated patients with COPD exacerbations experienced treatment failure after 3 months.58

COPD is a complex, heterogeneous, and progressive disease,⁵⁹ with myriad symptoms including airflow obstruction, mucus production, ciliary dysfunction, fibrosis, increased airway wall thickness, chronic bronchitis, fibrosis, and emphysema. Similar to SA and CRSwNP, COPD exacerbations can be brought about by multiple triggers, including infections, pollutants, cigarette smoke, and oxidative stress, leading to innate and adaptive immune pathway activation, inflammatory cell recruitment, cytokine/chemokine production, and elastase and matrix metalloproteinase activation.^{4,60}

The mechanisms behind COPD development have not been fully elucidated; however, epithelial cytokines, TSLP and IL-33 have been suggested to

AMJ

play a role in the underlying inflammatory processes associated with the disease.⁶¹ For example, TSLP is significantly increased in bronchoalveolar lavage fluid,⁶² epithelial tissue,⁶³ and airway smooth muscle⁶³ in patients with COPD compared with both smoking and non-smoking controls.⁶²⁻⁶⁴ There are also correlations of TSLP with significantly increased platelet counts and higher platelet aggregation rates.⁶⁵

Following bronchial epithelium injury, epithelial cytokine secretion, including TSLP, is increased, activating multiple cell types implicated in COPD pathophysiology, including T cells, ILC2s, and eosinophils.61 TSLP is also implicated in changes in non-immune system cells associated with COPD. For example, studies have shown higher expression of the TSLP receptor on fibrocytes from people with eosinophilic COPD compared with non-eosinophilic COPD,⁶⁶ and, in an *in vitro* study of human lung fibroblasts, TSLP upregulated collagen Type 1 and matrix metalloproteinase 1.⁶⁷ IL-33 can be released from the airway epithelium in response to COPDrelated triggers, such as cigarette smoke, microbes, and pollutants,² and compared with healthy controls, levels of IL-33 in exhaled breath condensate are significantly higher in patients with nonatopic COPD.68

Further, higher levels of IL-33 in plasma are significantly associated with an increased risk of COPD exacerbations, and intensity of IL-33 immunostaining in lung parenchyma is inversely correlated with the forced expiratory volume in 1 second in such patients.⁶⁹

Of note, cigarette smoking, a major risk factor for the development of COPD,⁵⁹ can also upregulate the expression of IL-33, with expression of this epithelial cytokine significantly correlated with smoking history.⁶⁹ However, levels of IL-33 in serum and sputum are significantly higher in patients with COPD compared with healthy non-smokers without COPD.⁶⁸ These findings, along with those of TSLP discussed above, suggest that, while smoking may increase epithelial cytokines independently of COPD, the levels in patients with this condition are above that of those caused only by cigarette smoke inhalation.

Conclusion

For SA, CRSwNP, and COPD, there are still a number of outstanding factors to explore

References

- Chipps BE et al. Number of patientreported asthma triggers predicts uncontrolled disease among specialisttreated patients with severe asthma. Ann Allergy Asthma Immunol. 2023;130(6):784-90.
- Calderon AA et al. Targeting interleukin-33 and thymic stromal lymphopoietin pathways for novel pulmonary therapeutics in asthma and COPD. Eur Respir Rev. 2023;32(167):220144.
- Laulajainen-Hongisto A et al. Airway epithelial dynamics in allergy and related chronic inflammatory airway diseases. Front Cell Dev Biol. 2020;8:204.
- Rabe KF et al. Targeting type 2 inflammation and epithelial alarmins in chronic obstructive pulmonary disease: a biologics outlook. Am J Respir Crit Care Med. 2023;208(4):395-405.
- Riera-Martínez L et al. The role of IL-33/ST2 in COPD and its future as an antibody therapy. Int J Mol Sci. 2023;24:8702.
- Yokota T et al. Isolation and characterization of a human interleukin cDNA clone, homologous to mouse B-cell stimulatory factor 1, that expresses B-cell- and T-cell-stimulating activities. Proc Natl Acad Sci U S A. 1986;83(16):5894-8.
- Azuma C et al. Cloning of cDNA for human T-cell replacing factor (interleukin-5) and comparison with the murine homologue. Nucleic Acids Res. 1986;14(22):9149-58.
- Minty A et al. Interleukin-13 is a new human lymphokine regulating inflammatory and immune responses. Nature. 1993;362(6417):248-50.

 Yang D, Oppenheim JJ. Antimicrobial proteins act as "alarmins" in joint immune defense. Arthritis Rheum. 2004;50(11):3401-3.

- Bachert C et al. The unified airway hypothesis: evidence from specific intervention with anti-IL-5 biologic therapy. J Allergy Clin Immunol Pract. 2023;11(9):2630-41.
- 11. Laidlaw TM et al. Chronic rhinosinusitis with nasal polyps and asthma. J Allergy Clin Immunol Pract. 2021;9(3):1133-41.
- Reche PA et al. Human thymic stromal lymphopoietin preferentially stimulates myeloid cells. J Immunol. 2001;167(1):336-43.
- Stier MT et al. Respiratory syncytial virus infection activates IL-13-producing group 2 innate lymphoid cells through thymic stromal lymphopoietin. J Allergy Clin Immunol. 2016;138(3):814-24.
- Lee J et al. IL-17E, a novel proinflammatory ligand for the IL-17 receptor homolog IL-17Rh1. J Biol Chem. 2001;276(2):1660-4.
- Baekkevold ES, et al. Molecular characterization of NF-HEV, a nuclear factor preferentially expressed in human high endothelial venules. Am J Pathol. 2003;163:69-79.
- Roan F et al. Epithelial cell-derived cytokines: more than just signaling the alarm. J Clin Invest. 2019;129(4):1441-51.
- 17. Gauvreau GM et al. Thymic stromal lymphopoietin: its role and potential as a therapeutic target in asthma. Expert Opin Ther Targets. 2020;24(8):777-92.
- Cayrol C, Girard JP. Interleukin-33 (IL-33): a nuclear cytokine from the IL-1 family. Immunol Rev. 2018;281(1):154-68.

when assessing disease pathogenesis and control. Airway epithelium plays a critical role in these conditions, with TSLP, IL-25, and IL-33 released following immune cell activation or epithelial damage. These cytokines can also be released by, and act on, a wide array of both immune and non-immune cells.² While TSLP, IL-25, and IL-33 have been linked to the pathogenesis of SA, CRSwNP, and COPD,^{15,16,20} further understanding of their role in these diseases is needed through both mechanistic and pre-clinical studies.

- Yao X et al. Interleukin (IL)-25: pleiotropic roles in asthma. Respirology. 2016;21(4):638-47.
- 20. Bartemes KR, Kita H. Dynamic role of epithelium-derived cytokines in asthma. Clin Immunol. 2012;143(3):222-35.
- 21. Porsbjerg CM et al. Anti-alarmins in asthma: targeting the airway epithelium with next-generation biologics. Eur Respir J. 2020;56(5):2000260.
- 22. Lambrecht BN, Hammad H. The immunology of asthma. Nat Immunol. 2015;16(1):45-56.
- 23. The Global Asthma Report 2022. Int J Tuberc Lung Dis. 2022;26 (Supp 1):1-104.
- 24. Calhoun WJ, Chupp GL. The new era of add-on asthma treatments: where do we stand? Allergy Asthma Clin Immunol. 2022;18(1):42.
- 25. Chen W et al. Economic burden of multimorbidity in patients with severe asthma: a 20-year population-based study. Thorax. 2019;74(12):1113-19.
- Denton E et al. Cluster analysis of inflammatory biomarker expression in the International Severe Asthma Registry. J Allergy Clin Immunol Pract. 2021;9(7):2680-8.
- Chen M et al. Overlap of allergic, eosinophilic and type 2 inflammatory subtypes in moderate-tosevere asthma. Clin Exp Allergy. 2021;51(4):546-55.
- Wang W et al. Bronchial Allergen challenge of patients with atopic asthma triggers an alarmin (IL-33, TSLP, and IL-25) response in the airways epithelium and submucosa. J Immunol. 2018;201(8):2221-31.

- 29. Cao L et al. TSLP promotes asthmatic airway remodeling via p38-STAT3 signaling pathway in human lung fibroblast. Exp Lung Res. 2018;44(6):288-301.
- Li Y et al. Elevated expression of IL-33 and TSLP in the airways of human asthmatics in vivo: a potential biomarker of severe refractory disease. J Immunol. 2018;200(7):2253-62.
- Shikotra A et al. Increased expression of immunoreactive thymic stromal lymphopoietin in patients with severe asthma. J Allergy Clin Immunol. 2012;129(1):104-11.
- Ko HK et al. Blood tryptase and thymic stromal lymphopoietin levels predict the risk of exacerbation in severe asthma. Sci Rep. 2021;11(1):8425.
- Bae CH et al. Effect of thymic stromal lymphopoietin on MUC5B expression in human airway epithelial cells. Biochem Biophys Res Commun. 2014;448(2):2315.
- Schleimer RP. Immunopathogenesis of chronic rhinosinusitis and nasal polyposis. Annu Rev Pathol. 2017;12:331-57.
- 35. Chen S et al. Systematic literature review of the epidemiology and clinical burden of chronic rhinosinusitis with nasal polyposis. Curr Med Res Opin. 2020;36(11):1897-1911.
- Khan A et al. The GALEN rhinosinusitis cohort: chronic rhinosinusitis with nasal polyps affects health-related quality of life. Rhinology. 2019;57(5):343-51.
- Bhattacharyya N et al. Cost burden and resource utilization in patients with chronic rhinosinusitis and nasal polyps. Laryngoscope. 2019;129(9):1969-75.
- Claeys N et al. Patients unmet needs in chronic rhinosinusitis with nasal polyps care: a patient advisory board statement of EUFOREA. Front Allergy. 2021;2:761388.
- Mullol J et al. Chronic rhinosinusitis with nasal polyps: quality of life in the biologics era. J Allergy Clin Immunol Pract. 2022;10(6):1434-53.
- 40. Braunstahl GJ et al. Nasal allergen provocation induces adhesion molecule expression and tissue eosinophilia in upper and lower airways. J Allergy Clin Immunol. 2001;107(3):469-76.
- Chiarella E et al. Nasal polyposis: insights in epithelial-mesenchymal transition and differentiation of polyp mesenchymal stem cells. Int J Mol Sci. 2020;21(18):6878.
- 42. Kato A et al. Mechanisms and pathogenesis of chronic rhinosinusitis.

J Allergy Clin Immunol. 2022;149(5):1491-1503.

- 43. Stevens WW et al. Associations between inflammatory endotypes and clinical presentations in chronic rhinosinusitis. J Allergy Clin Immunol Pract. 2019;7(8):2812-20.
- 44. Staudacher AG et al. Use of endotypes, phenotypes, and inflammatory markers to guide treatment decisions in chronic rhinosinusitis. Ann Allergy Asthma Immunol. 2020;124(4):318-25.
- Hao D et al. An integrated analysis of inflammatory endotypes and clinical characteristics in chronic rhinosinusitis with nasal polyps. J Inflamm Res. 2022;15:5557-65.
- Kobayashi Y et al. Reduced local response to corticosteroids in eosinophilic chronic rhinosinusitis with asthma. Biomolecules. 2020;10(2):326.
- 47. Liu R et al. Elevated microRNA-21 is a brake of inflammation involved in the development of nasal polyps. Front Immunol. 2021;12:530488.
- Kim DK et al. The role of interleukin-33 in chronic rhinosinusitis. Thorax. 2017;72(7):635-45.
- 49. van der Ploeg EK et al. Steroid-resistant human inflammatory ILC2s are marked by CD45RO and elevated in type 2 respiratory diseases. Sci Immunol. 2021;6(55):eabd3489.
- Xue JM et al. Livin in synergy with Ras induces and sustains corticosteroid resistance in the airway mucosa. Int J Biol Sci. 2021;17(8):2089-98.
- Zhang M et al. Hypoxia induces the production of epithelial-derived cytokines in eosinophilic chronic rhinosinusitis with nasal polyps. Int Immunopharmacol. 2023;121:110559.
- 52. Liao B et al. Interaction of thymic stromal lymphopoietin, IL-33, and their receptors in epithelial cells in eosinophilic chronic rhinosinusitis with nasal polyps. Allergy. 2015;70(9):1169-80.
- 53. Adeloye D et al. Global, regional, and national prevalence of, and risk factors for, chronic obstructive pulmonary disease (COPD) in 2019: a systematic review and modelling analysis. Lancet Respir Med. 2022;10(5):447-58.
- 54. Mathers CD, Loncar D. Projections of Global Mortality and Burden of Disease from 2002 to 2030. PLoS Med. 2006;3(11):e442
- 55. Chen S et al. The global economic burden of chronic obstructive pulmonary disease for 204 countries

and territories in 2020-50: a health-augmented macroeconomic modelling study. Lancet Glob Health. 2023;11(8):e1183-93.

- 56. Liu Y et al. Trends in the prevalence of chronic obstructive pulmonary disease among adults aged ≥18 years - United States, 2011-2021. MMWR Morb Mortal Wkly Rep. 2023;72(46):1250-6.
- 57. American Lung Association. COPD trends brief: Mortality. 2024. Available at: https://www.lung.org/research/ trends-in-lung-disease/copd-trendsbrief/copd-mortality. Last accessed: 25 September 2024.
- Mathioudakis AG et al. Assessing treatment success or failure as an outcome in randomised clinical trials of COPD exacerbations. A Meta-Epidemiological Study. Biomedicines. 2021;9(12):1837.
- 59. Global Initiative for Chronic Obstructive Lung Disease. Global strategy for the diagnosis, managment, and prevention of chronic obstructive pulmonary disease. Available at: https://goldcopd. org/wp-content/uploads/2024/02/ GOLD-2024_v1.2-11Jan24_WMV.pdf. Published 2024. Last accessed: 25 September 2024.
- 60. Alfahad AJ et al. Current views in chronic obstructive pulmonary disease pathogenesis and management. Saudi Pharm J. 2021;29(12):1361-73.
- 61. Barnes PJ. Inflammatory endotypes in COPD. Allergy. 2019;74(7):1249-56.
- 62. Ying S et al. Expression and cellular provenance of thymic stromal lymphopoietin and chemokines in patients with severe asthma and chronic obstructive pulmonary disease. J Immunol. 2008;181(4):2790-8.
- 63. Paplinska-Goryca M et al. Epithelialmacrophage-dendritic cell interactions impact alarmins expression in asthma and COPD. Clin Immunol. 2020;215:108421.
- 64. Zhang K et al. Constitutive and inducible thymic stromal lymphopoietin expression in human airway smooth muscle cells: role in chronic obstructive pulmonary disease. Am J Physiol Lung Cell Mol Physiol. 2007;293(2):L375-82.
- 65. Wu L et al. Effect of TSLP on the function of platelets and IL-25 in chronic obstructive pulmonary disease. Int J Clin Exp Med. 2019;12:4942-8.
- 66. Tworek D et al. Epithelial alarmin expression on circulating fibrocytes in stable, non-atopic, eosinophilic COPD. Chest. 2020;157(6):A281.